

## **ACTION OF POLYPEPTIDE GROWTH FACTORS IN COLON CANCER; DEVELOPMENT OF NEW THERAPEUTIC APPROACHES**

**Subhas Chakrabarty, Sharon Reynolds, Hong mei Wang and Sriram Rajagopal**

*Division of Laboratory Medicine, University of Texas, M.D. Anderson Cancer Center, Houston, TX 77030*

### **TABLE OF CONTENTS**

1. Abstract
2. Introduction
3. Discussion
  - 3.1. Adhesion molecules and TGF-beta
  - 3.2. Mechanisms of action of TGF-beta in colon cancer, role of protein kinase C
  - 3.3. PKC alpha-a potential therapeutic target
  - 3.4. EGF family of growth factors and the potential of EGF receptor as a therapeutic target
  - 3.5. Other potential therapeutic targets
  - 3.6. Therapeutic approach
  - 3.7. Basis of selectivity against colon cancer
  - 3.8. Perspectives
4. Acknowledgments
5. References

### **1. ABSTRACT**

There has been no significant improvement in the treatment of metastatic colon cancer over the last four decades. A major reason for therapeutic failure is the nonselective nature of conventional chemotherapy. Therefore, new selective therapeutic approaches must be explored to improve survival. Recent advances in cell and molecular biology have opened up new avenues of developing selective molecular therapy for colon cancer. In view of the heterogeneity of cells in colon cancer, it is likely that the use of more than one cellular target will be required to achieve a significant antitumor response. Mechanistic studies of how proliferation, differentiation and malignant properties are controlled by negative growth factors, positive growth factors and adhesion molecules have allowed the identification of several molecular targets of attack in colon cancer. Disruption of one or more of these targets should have a highly antiproliferative and/or cytotoxic effect on colon cancer cells. Additionally, disrupting the expression of these targets may augment sensitivity to conventional chemotherapy. In this article, we will discuss how polypeptide growth factors act in colon cancer cells, identify several molecular targets of attack and discuss strategies for selective disruption of these targets in colon cancer. Where appropriate, the biologic similarities or differences by comparison with other types of tumor will also be discussed.

### **2. INTRODUCTION**

Metastatic human colon cancer is a highly refractory malignant disease which afflicts both men and women in equal proportion. The antimetabolite, 5-fluorouracil (5-FU), used to treat colon cancer over 4 decades ago, is still the drug of choice today for the treatment of metastatic disease. Despite some improvements in adjuvant therapy using 5-FU in combination with leucovorin or levamisole, almost half of all colorectal cancer patients will die of metastatic disease (1). Therefore, new therapeutic approaches must be explored to improve survival. An understanding of the biology behind malignant transformation and the mechanisms sustaining the transformed phenotype may provide avenues of therapeutic intervention.

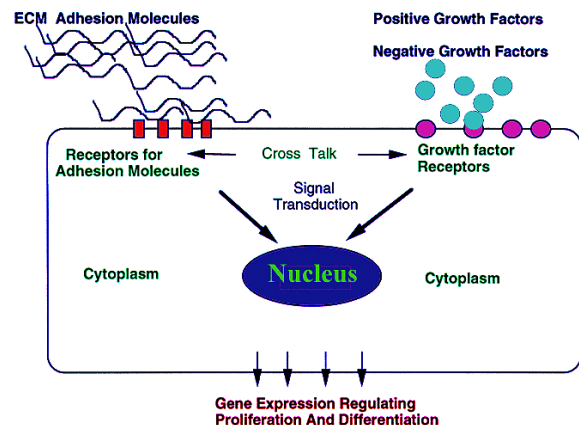
Control of cellular proliferation, differentiation and adhesion are complex biological processes in which different biological systems interact to achieve control. Cell-matrix and cell-cell adhesion are important cellular processes in regulating cellular proliferation and differentiation (2-5). Extracellular matrix (ECM) adhesion molecules modulate gene expression in epithelial cells in a tissue specific manner (5-7). Disruption of these processes is a hallmark of malignant transformation and plays a critical role in tumor progression and the behavior of malignant cells (8-10).

Polypeptide growth factors constitute a potent class of extracellular and/or intracellular signal molecules in regulating cellular proliferation and differentiation (11-13). Aberrant expression of growth factors and/or aberrant responses to growth factors may circumvent the normal pathway of differentiation, leading to cellular transformation, tumor progression and maintenance of the transformed phenotype (11, 14). Transforming growth factor (TGF) beta constitutes a class of multi-functional

---

Received 9/3/97 Accepted 9/15/97

Send correspondence to: Dr. Subhas Chakrabarty, Division of Laboratory Medicine, Box 73, University of Texas, M.D. Anderson Cancer Center, 1515 Holcombe Boulevard, Houston, TX 77030, Tel: (713) 792-5538, Fax: (713) 794-5765, E-mail: schakrab@notes.mdacc.tmc.



**Figure 1.** Interaction of ECM adhesion molecules with growth factors

polypeptide growth factors that regulate proliferation and differentiation in many cell types (15-17) and suppress malignancy in TGF-beta-responsive epithelial cancer cells (18-20). Loss of responsiveness to TGF-beta is thought to be a mechanism of escape from normal growth control in malignant cells (21-22). The ability of TGF-beta to induce a more benign and differentiated phenotype in malignant cells is partly attributable to its ability to upregulate the synthesis of ECM adhesion molecules (23-28). Thus, TGF-beta may be viewed as a class of negative growth factors.

The epidermal growth factor (EGF) family of polypeptide growth factors such as EGF and TGF-alpha, on the other hand, are potent stimulators of cellular proliferation and stimulate the malignant behavior of many epithelial-derived cancer cells (29-35). Thus, the EGF family of growth factors may be viewed as a class of positive growth factors. How ECM adhesion molecules and growth factors interact in controlling proliferation, differentiation and adhesion is depicted in Figure 1.

In this review, we will discuss how negative and positive growth factors act in colon cancer and review the rationale and strategy behind the development of new therapeutic approaches. Where appropriate, the biologic similarities or differences by comparison with other tumor types or other agents involved in regulating differentiation will also be discussed.

### 3. DISCUSSION

#### 3.1. Adhesion molecules and TGF-beta

Fibronectin (FN) is a transformation-sensitive ECM adhesion molecule. The expression of FN in many transformed cells is lost (36-40). FN interacts with specific cell-surface FN receptor to initiate intracellular signal transduction leading to the regulation of gene expression (41-45). A major species of FN receptor is the integrin alpha5beta1 which has been shown to act as a tumor suppressor (46-47). Interestingly, when transformed cells are treated with differentiation-inducing agents, these agents induce a higher level of FN and FN receptor

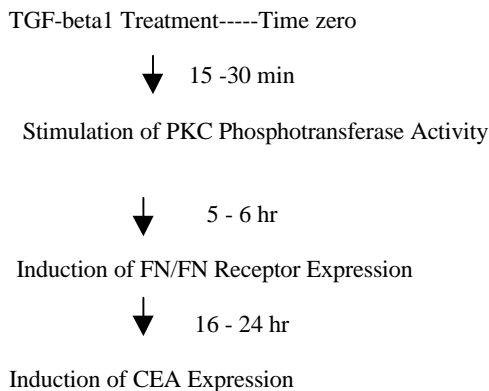
expression in conjunction with the restoration of a more normal or benign phenotype to the transformed cells (36-38). In normal and malignant cell hybrids, the malignant phenotype is suppressed and the hybrids express a high level of FN by comparison with the malignant cells (48). However, blockade of FN synthesis in the hybrids abrogates the suppression of the malignant phenotype (48). The expression of FN and FN receptor is tightly regulated in the process of transformation and the induction of differentiation of the transformed cells. When mouse embryonic fibroblasts are transformed with a chemical carcinogen, the expression of FN and FN receptor is significantly reduced (36-38). Induction of differentiation of the transformed cells by treatment with differentiation-inducing agents restores normal growth control and cellular morphology to the transformed cells with a concurrent restoration of the expression of FN and FN receptor (36-38). Blockade of FN synthesis in the transformed cells downregulates the ability of differentiation-inducing agents to induce FN and FN receptor expression and circumvents the induction of normal cellular morphology (49). Thus, both FN and FN receptor play a functional role in chemical transformation and the induction of differentiation in transformed cells.

In this context, human colon cancer cells produce very little FN (50). The production of FN, however, is upregulated when colon cancer cells are induced to differentiate towards a more normal or benign phenotype by treatment with TGF-beta or other differentiation-inducing chemicals (23-28). Patients with FN-positive invasive breast carcinomas fare significantly better than patients with FN-negative tumors in terms of relapse-free survival. In addition, the FN staining pattern is independently correlated with relapse-free survival and nodal status (51).

Interestingly, contrary to the colon and breast tumor types, both androgen-responsive and non-responsive prostate cancer cells produce abundant amounts of FN (52-53). The biological role of these molecules in the malignant phenotype of prostate cancer cells is not known. However, it is now known that the ECM and stroma play a critical role in malignant transformation and progression of the prostatic epithelium (54). It is of interest to note that (contrary to colon and breast cancer) a high level of FN expression has been reported in prostate cancer tissue by comparison with normal prostate and hyperplastic tissues (55-56).

Three isoforms of TGF-beta (TGF-beta1, TGF-beta2 and TGF-beta3) are expressed in mammals and overall, the biological activities of these isoforms are very similar *in vitro* (57). Therefore, for the purpose of discussion, the generic term TGF-beta will be used in this article. TGF-beta interacts with a family of high-affinity cell-surface receptors through which it mediates its biological action. Most cell types, including human colon cancer cells possess such receptors for TGF-beta (57-60). The family of TGF-beta receptors interact to mediate the action of TGF-beta (61-65). The post-receptor or intracellular mechanisms through which

## Growth factors and colon cancer



**Figure 2.** Intracellular events following TGF-beta binding to human colon cancer cells

TGF-beta acts, however, are poorly understood. Intricate and complex mechanisms appear to be involved in the action of TGF-beta. It has been proposed that multiple TGF-beta signal-transducing pathways exist (66). Both differences and similarities are likely to exist when the biological action of TGF-beta and its mechanisms of action are compared in different cell types (15, 66-72).

TGF-beta elicits diverse responses in TGF-beta-responsive human colon cancer cells. These responses include inhibition of proliferation, invasive capability, growth in soft agarose and tumor formation in athymic mice (18-20, 25, 73). At the molecular level, TGF-beta upregulates the expression of the adhesion molecules, FN and laminin (LM), the cell-surface receptors for these molecules, and the intercellular adhesion molecule, carcinoembryonic antigen (CEA) (23-28). In addition, TGF-beta upregulates the expression of epithelial-associated cytokeratins, reorganizes the cytoskeleton and induces increased adhesion and spread on the ECM (23-28). The net effect of TGF-beta action is the induction of a more differentiated or benign phenotype in colon cancer cells. Thus, how TGF-beta controls these cellular responses is of great interest to cancer biologists and such knowledge may provide leads in the development of novel therapeutics against colon cancer. It is hypothesized that the development of resistance to TGF-beta is a mechanism of tumor progression and escape from normal growth and differentiation control. Indeed, mutated TGF-beta type II receptor has been found to be associated with highly malignant TGF-beta-non-responsive human colon cancer cells (22) and suppression of TGF-beta autocrine activity promotes malignancy (18). The loss of responsiveness to TGF-beta may also be an important mechanism of escape from normal growth control in prostate and breast cancers (21, 74-75).

### 3.2. Mechanisms of action of TGF-beta in colon cancer, role of protein kinase C

Protein kinase C (PKC) is an ubiquitous calcium- and phospholipid-dependent protein kinase involved in transmembrane signal transduction (76). It may also play a

role in growth control and carcinogenesis in the colon (77-79). At least 11 PKC isoforms have been identified and characterized (79-81). The functional significance of individual isoforms, however, is not well understood. TGF-beta stimulates a rapid rise in PKC phosphotransferase activity (69) and the kinetics of events that follow are depicted in Figure 2.

Earlier work has shown that chemical inhibitors of PKC were able to block the ability of TGF-beta to induce CEA expression, suggesting the functional role of this enzyme in the signal pathway of TGF-beta leading to the induction of CEA (69). Interestingly, blockade of FN induction (an earlier event) by expressing antisense FN RNA in the cells also blocked the induction of CEA, suggesting a functional role of FN in the induction of CEA (28). Exactly how FN interacts with the TGF-beta pathway leading to the induction of CEA is not known. FN can interact with other components of the ECM in regulating gene expression (82) or newly synthesized FN may act through a feed-back mechanism, interacting with the FN receptor and initiating signal-transduction cascades (41-45, 83) that are required for TGF-beta to mediate its action on the CEA gene.

Like the mouse embryonic fibroblasts described above, the expression of FN and FN receptor is tightly regulated in human colon cancer cells. Treatment of colon cancer cells with TGF-beta upmodulates the expression of both FN and FN receptor (27). Blockade of FN synthesis by the cellular expression of antisense FN RNA circumvented the ability of TGF-beta to upregulate the expression of both FN and its receptor (84). This makes sense from a cellular physiologic point of view. If the cells were to respond to the physiologic effect of an increased level of FN expression, the level of receptor expression needs to increase to accommodate the ligand. The mechanisms behind the coregulation of the FN ligand and its receptor by TGF-beta remain to be elucidated.

Recent work identifies PKCalpha isoform in controlling the adhesion response (induction of adhesion molecules and receptors for these molecules) to TGF-beta. Expression of antisense PKCalpha RNA (by transfection with an antisense PKCalpha expression vector) in human colon cancer cells downregulated PKCalpha protein expression and attenuated the ability of TGF-beta to upregulate the expression of FN, FN receptor and CEA (70). Interestingly, perturbation of PKCalpha did not interfere with the molecular pathway of TGF-beta leading to the inhibition of cellular proliferation (unpublished observation). These results suggest that PKCalpha is a focal point in controlling the adhesion signal pathway of TGF-beta but not its antiproliferative pathways. Further work is in progress to confirm the validity of this hypothesis.

We wish to point out that the induction of FN and CEA is not a unique property of TGF-beta. Many differentiation-inducing agents such as retinoic acid, difluoromethylornithine, dimethylformamide, sodium butyrate and sodium suramin will inhibit the malignant properties of human colon cancer cells with a concurrent

## Growth factors and colon cancer

upregulation of FN and CEA synthesis (85). The mechanisms of action of these differentiation-inducing chemicals are probably quite different from that of TGF-beta, and the differentiation-inducing capability of these chemicals varies and is contingent upon the phenotype of the target cells (85). This is not surprising in view of the phenotypic heterogeneity of colon cancer cells.

### 3.3. PKC alpha - a potential therapeutic target

Alterations in PKC expression have been suggested to play a role in tumor progression and in the maintenance of the malignant phenotype (76-77, 86-89). In addition, PKC is implicated to play a functional role in multi-drug resistance (90-91). Specifically, PKC $\alpha$  activates the MDR-1 gene product, gp170, by phosphorylation and thus increases efflux of drugs from the cell (92). In the colonic epithelium, continuous activation of PKC by unsaturated diacylglycerols in the intestine may be responsible for intrinsic drug resistance (93). PKC $\alpha$  isoform is implicated to play a role in drug resistance in colon cancer cells (94). It has recently been shown that PKC $\alpha$  can be targeted to potentiate the responses of colon cancer cells to anticancer drugs. Expression of PKC $\alpha$  antisense RNA (but not beta or gamma isoforms) in human colon cancer cells resulted in down-modulation of PKC expression and increased the cytotoxic effect of 5-fluorouracil, mitomycin C and vincristine by several folds (95). Therefore, PKC $\alpha$  is a good target for the purpose of potentiating sensitivity to conventional anticancer drugs.

### 3.4. EGF family of growth factors and the potential of EGF receptor as a therapeutic target

Human colon cancer cells produce and respond to the EGF related family of polypeptide growth factors (32-33, 73, 96). The EGF family of growth factors consists of EGF, TGF- $\alpha$ , cripto and amphiregulin (29-31). These growth factors are potent stimulators of cellular proliferation, and overexpression of one or more of these growth factors or receptors for these factors have been implicated to play a causal role in the transformation and maintenance of the transformed phenotype (11-12, 14, 35, 97-102). EGF related growth factors mediate their action through a common receptor, the EGF receptor (EGFR) (29). Stimulation of the EGFR by growth factors induces proliferation and malignant cell behavior (e.g. propensity to grow in semi-solid medium and invasion of matrigel (32-34, 101)). In addition, highly metastatic human colon cancer cells express a relatively high level of EGFR (35).

Cells can respond to these growth factors via an autocrine mode (i.e. the cells secrete these factors and the factors then bind to the cell-surface EGFR thus, activating the EGFR); a paracrine mode (i.e. cells respond to factors produced by other cells via the cell-surface EGFR); or an intracrine mode (i.e. cells produce and respond to these factors inside the cells, bypassing the cell-surface EGFR) (12-13). Previous work shows that human colon cancer cell lines, including some highly aggressive lines, produce and secrete both EGF and TGF. Some cell lines utilize these growth factors via the cell-surface EGFR (i.e. susceptible to blockade by anti-EGFR antibodies), others do not (32) i.e. these cells may utilize growth factors via an intracrine

mode. Subsequent work supported the hypothesis that these cells utilize growth factors in an intracrine manner because the intracellular expression of antisense EGFR RNA in these cells inhibited the proliferation and malignant behavior of these cells (33-34). Some human colon cancer cell lines are very sensitive to EGFR blockade by the intracellular expression of antisense EGFR RNA through transfection with an antisense EGFR expression vector (33-34), while others are relatively resistant. The expression of antisense EGFR RNA in the sensitive cells results in a significant downmodulation of EGFR protein and mRNA expression with a concurrent inhibition of cellular proliferation and malignant properties (33-34). Therefore, EGFR may be a good molecular target to be used against colon cancer.

The EGFR may also serve as a good target of attack for other epithelial cancer. Studies using human prostate carcinoma tissues and prostate carcinoma cell lines strongly implicate the participation of EGF-related molecules and EGF receptor in the pathogenesis of prostate cancer (103-104). In fact, the growth stimulatory effect of androgen in androgen-responsive prostate cancer cells is attributable to the EGF/TGF- $\alpha$ -EGF receptor loop (105-106) and EGF can activate the androgen-signaling chain in the absence of androgens. This loop also participates in the cellular proliferation of androgen-independent prostate cancer cells (107). In breast carcinoma, the expression of Her-2/neu, a member of the EGFR family of receptors, is significantly elevated and correlates with poor prognosis, tumor progression and drug resistance (108-109). EGF and TGF- $\alpha$  also act as positive regulators in breast cancer (110-111). Thus, Her-2/neu and EGFR are also good therapeutic targets for breast cancer.

### 3.5. Other potential therapeutic targets

Cyclin dependent kinase 2 (CDK2) is an intracellular signal-transduction molecule that functions in regulating cell cycle progression (112). It initiates cell cycle progression by phosphorylating specific cellular target molecules (112). Many growth factors mediate their effect through CDK2. Blockade of EGFR with anti-EGFR antibody blocks CDK2 activity and results in the induction of G1 arrest (113-114). Thus, disrupting CDK2 and EGFR may lead to a "double whammy" effect on the cancer cells: choking off the action of the EGF related family of growth factors and simultaneously blocking the action of CDK2, the activity of which is required for cell cycle progression. In addition, since CDK2 is a downstream target of the EGFR, cells that are resistant to EGFR blockade may have developed mechanisms of escape from CDK2 inactivation. Therefore, it is likely that cells that are resistant to EGFR blockade will be susceptible to the direct disruption of CDK2.

As discussed above, TGF-beta suppresses malignant tumor growth and induces differentiation in human colon cancer cells. Escape from TGF-beta control, through lack of TGF-beta receptor expression or expression of mutated receptor, is thought to play an important role in the tumor progression and malignant growth of colon

## Growth factors and colon cancer

cancer (22). An important postreceptor target of TGF-beta is also CDK2. TGF-beta induces G1 cell cycle arrest by inhibiting the activity of CDK2 (115-116). Thus, disrupting CDK2 may mimic the effect of TGF-beta and furthermore it may also mimic TGF-beta effect in TGF-beta-resistant cells because of direct disruption of a downstream postreceptor target of TGF-beta. Thus, disruption of CDK2 and EGFR together may deliver a "triple whammy" effect to the cancer cells.

As discussed above, cell-matrix and cell-cell interactions play critical roles in controlling the normal growth and differentiation programs of epithelial cells (2-7) and that abnormal cell-matrix and/or cell-cell interactions is a hallmark of malignant transformation (8-11, 36-38). Interactions of extracellular matrix components with specific cell-surface receptors turn on intracellular signal transduction mechanisms leading to the control of gene expression in a tissue-specific manner (41-45). Focal adhesion kinase (FAK) is an intracellular signaling molecule (tyrosine protein kinase) associated with the intracellular tails of cell-surface integrin receptors that bind to extracellular matrix components such as fibronectin (117-119). When normal epithelial cells are detached from the matrix they undergo programmed cell death (120). Activation of FAK allows epithelial cells to become immortalized and undergo malignant transformation (120). FAK has been shown to be overexpressed in colon cancer (121) and disruption of FAK expression by antisense FAK oligonucleotides is cytotoxic to highly malignant human rhabdomyosarcoma cells in culture (122). Therefore, we hypothesize that disrupting FAK expression will lead to growth inhibition and/or cell death in colon cancer.

### 3.6. Therapeutic Strategies

In view of the heterogeneity of colon cancer cells, and the many mechanisms that cancer cells use to circumvent normal cellular control, it is likely that a multi-targeting and multi-modal type of approach is required to achieve a significant antitumor response. We have discussed at least 4 molecular targets of attack in colon cancer. Some of these targets, (e.g. EGFR and FAK) though overexpressed in colon cancer, are also expressed in most normal tissues. Therefore, success is more likely, if a basis for selective disruption of these targets in colon cancer cells exists in the therapeutic design. In addition, disrupting the expression of these targets (a good example is PKC alpha) may augment sensitivity to conventional chemotherapy. Thus, the molecular approach has the potential to be used in a truly multi-targeting and multi-modal manner with conventional chemotherapy to induce a high level of antitumor response.

Over the last several years this laboratory has used antisense mammalian expression vectors, under the control of viral promoters, for the purpose of disrupting the expression and/or function of specific cellular molecules. To this end, we have been successful in disrupting the expression and function of adhesion molecules such as fibronectin (28, 84) and other important signal molecules such as protein kinase C isoform (70, 95) and EGFR (33-34). In each of these cases, we have demonstrated that

following transfection with the antisense expression vectors, the cells expressed the antisense RNA species and that the effect of antisense RNA expression resulted in a downregulation of the corresponding mRNA and protein expression. Exactly how the antisense RNA work inside the cells in disrupting protein expression is not known. It is likely that the expression of the antisense strand RNA interferes with translation of the sense mRNA strand and/or the antisense strand binds to its complimentary sense strand leading to rapid degradation of the sense mRNA (123). Therefore, we propose to use this approach for the purpose of disrupting the expression of key molecular targets in colon cancer.

### 3.7. Basis of selectivity against colon cancer

CEA is preferentially expressed in colon cancer cells and other tumors of the gastrointestinal tract (124). CEA is also expressed, albeit at a low level, in the normal gastrointestinal tracts (125-126). CEA, however, is not expressed in the majority of normal tissues including the bone marrow, heart, kidney and liver (125-126) (the liver being the major site of metastasis for colon cancer). The CEA promoter has been shown recently to be capable of driving the expression of genes only in CEA-producing cancer cells (127-128); i.e. only cells expressing CEA have the capability, or possess the transcription initiation factors necessary for turning on the CEA promoter. Therefore, if the antisense expression vectors targeting EGFR, CDK2, FAK or PKC were constructed under the control of the CEA promoter, there is a good chance of achieving a good degree of selectivity against colon cancer cells. One may envision infusion of these "DNA drugs" which are selectively active in colon cancer even though they might be taken up by most normal tissues and may eventually be degraded.

### 3.8. Perspective

Recent advances in cell and molecular biology have opened up new avenues of therapeutic approaches in attacking colon cancer. Chances of success are much improved with a multi-modal approach. A multi-modal approach is particularly attractive if a basis of selectivity exists against colon cancer cells. In the context of the "antisense DNA drugs", this is only an embryonic beginning and much research needs to be performed to determine their efficacy and optimal administrative regimens. A key to success lies in the development of non-toxic drug delivery systems for systemic therapy or regional therapy of liver metastasis. The use of non-toxic cationic liposomes (129-131) for drug delivery is quite attractive and may allow frequent infusion of "DNA drugs" as needed to achieve a favorable antitumor response.

## 4. ACKNOWLEDGMENTS

Supported by USPHS grant R01CA47775 and research grants from the Elsa U Pardee Foundation and the Physician Referral Service of M.D. Anderson Cancer Center.

### 5. REFERENCES

1. P.G. Johnston & C.J. Allegra: Colorectal cancer biology: clinical implications. *Seminars in Oncology* 22, 418-432 (1995)
2. E.D. Hay: Extracellular Matrix. *J Cell Biol* 91, 205s-223s (1981)
3. G.M. Edelman: Cell adhesion molecules. *Science* (Washington, D.C.) 219, 450-457 (1983)
4. E. Ruoslahti & M.D. Pierschbacher: New perspectives in cell adhesion: RGD and integrins. *Science* (Washington, D.C.) 238, 491-497 (1987)
5. C.Q. Lin & M.J. Bissel: Multi-faceted regulation of cell differentiation by extracellular matrix. *FASEB J* 7, 737-741 (1993)
6. C.D. Roskelley, P.Y. Desprez & M.J. Bissell: Extracellular matrix-dependent tissue-specific gene expression in mammary epithelial cells required both physical and biochemical signal transduction. *Proc Natl Acad Sci* 91, 12378-12382 (1994)
7. E.A. Clark & J.S. Brugge: Integrins and signal transduction pathway: the road taken. *Science* (Washington, D.C.) 268, 233-238 (1995)
8. R.H. Kramer, J. Enestein, D.M. Ramos, M.P. Vu & Y.F. Cheng: 1991. The role of integrin receptors in tumor cell adhesion to the microvasculature. In: *Microcirculation in cancer metastasis*. Eds: Buchanan M R, Weis L, CRC Press, Boca Raton (1991)
9. R.L. Juliano: Membrane receptors for extracellular matrix macromolecules: relationship to cell adhesion and tumor metastasis. *Biochim Biophys Acta* 907, 261-278 (1987)
10. R.L. Juliano: Signal transduction by integrins and its role in the regulation of tumor growth. *Cancer Met Rev* 13, 25-30 (1994)
11. Cross, M. and Dexter, T.M. Growth factors in development, transformation, and tumorigenesis. *Cell* 64:271-280, 1991.
11. M. Cross & T.M. Dexter: Growth factors in development, transformation and tumorigenesis. *Cell* 64, 271-280 (1991)
12. T.M. Browder, C.E. Dunbar & A.W. Nienhuis: Private and public autocrine loops in neoplastic cells. *Cancer Cells* 1,9-17 (1989)
13. A. Logan: Intracrine regulation at the nucleus - a further mechanism of growth factor activity? *J Endocrinol* 125, 339-343 (1990)
14. S. Kerbel: Growth factors as mediators of malignant tumor progression. *Cancer Metastasis Rev* 12, 215-217 (1993)
15. H.L. Moses, E.Y. Yang & J.A. Pietsenpol: TGF $\beta$  stimulation and inhibition of cell proliferation: new mechanistic insights. *Cell* 63, 245-247 (1990).
16. J.A. Barnard, R.M. Lyons & H.L. Moses: The cell biology of transforming growth factor  $\beta$ . *Biochimica Biophysica Acta* 1032, 79-87 (1990)
17. S. Chakrabarty, D. Fan & J. Varani: Regulation of differentiation and cellular proliferation in human colon carcinoma cells by transforming growth factor beta 1 and beta 2. *Int J Cancer* 46, 493-499 (1990)
18. S.P. Wu, L-Z. Sun, L-Z, J.K.V. Willson, L. Humphrey, R. Kerbel & M.G. Brattain: Repression of autocrine transforming growth factor beta1 and beta2 in quiescent CBS colon carcinoma cells leads to progression of tumorigenic properties. *Cell Growth and Diff* 4, 115-123 (1993)
19. L. Sun, S.P. Wu, K. Coleman, K.C. Fields, L.E. Humphrey & M.G. Brattain: Autocrine transforming growth factor-beta1 and beta2 expression is increased by cell crowding and quiescence in colon carcinoma cells. *Exp Cell Res* 214, 215-224 (1994)
20. S. Wu, D. Theodorescu, R.S. Kerbel, J.K.V. Willson, K.M. Mulder, L.E. Humphrey & M.G. Brattain: TGF-beta 1 is an autocrine-negative growth regulator of human colon carcinoma FET cells in vivo as revealed by transfection of an antisense expression vector. *J Cell Biol* 116, 187-196 (1992)
21. M.S. Steiner: Transforming growth factor-beta and prostate cancer. *World J Urol* 13, 329-336 (1995)
22. S. Markowitz, J. Wang, L. Myeroff, R. Parsons, L-Z Sun, J. Lutterbaugh, R.S. Fan, E. Zborowska, B. Vogelstein, M.G. Brattain & J.K.V. Willson: Inactivation of the type II TGF-beta receptor in colon cancer cells with microsatellite instability. *Science* (Washington, D.C.) 268, 1336-1338 (1995)
23. S. Chakrabarty, A. Tobon, J. Varani & M.G. Brattain: Induction of carcinoembryonic antigen secretion and modulation of protein secretion/expression and fibronectin/laminin expression in human colon carcinoma cells by transforming growth factor-beta. *Cancer Res* 48, 4059-4064 (1988)
24. S. Chakrabarty, Y. Jan, M.G. Brattain, A. Tobon & J. Varani: Diverse cellular responses elicited from human colon carcinoma cells by transforming growth factor- $\beta$ . *Cancer Res* 49, 2112-2117 (1989)
25. S. Chakrabarty, D. Fan & J. Varani: Regulation of

## Growth factors and colon cancer

differentiation and cellular proliferation in human colon carcinoma cells by transforming growth factor beta 1 and beta 2. *Int J Cancer* 46, 493-499 (1990)

26. S. Chakrabarty: Regulation of human colon carcinoma cell adhesion to extracellular matrix by transforming growth factor-beta1. *Int J Cancer* 50, 968-973 (1992)

27. S. Huang & S. Chakrabarty: Regulation of fibronectin and laminin receptor expression, fibronectin and laminin secretion in human colon cancer cells by transforming growth factor beta1. *Int J Cancer* 57, 742-746 (1994)

28. S. Huang & S. Chakrabarty: Expression of antisense fibronectin RNA in human colon carcinoma cells disrupts the regulation of carcinoembryonic antigen by transforming growth factor beta1. *J Biol Chem* 269, 28764-28768 (1994)

29. R.W. Donaldson, S. Nishibe, & G. Carpenter: The structure and physiology of epidermal growth factor and its receptor. In: *Advances in Growth Hormone and Growth Factor Research*. Eds: Miller E.E, Cocchi D, Locatelli V, Pythagora Press, Roma-Milano, Springer Verlag, Berlin-Heidelberg (1989)

30. M. Shoyab, G.D. Plowman, V.L. McDonald, J.G. Bradley & G.J. Todaro: Structure and function of human amphiregulin: a member of the epidermal growth factor family. *Science* (Washington D.C.) 243, 1074-1076 (1989)

31. F. Ciardiello, G. Tortora, C. Bianco, M.P. Selvam, F. Basolo, G. Fontanini, F. Pacifico, N. Normanno, R. Brandt, M.G. Perisco, D.S. Salomon & R. Bianco: Inhibition of CRIPTO expression and tumorigenicity in human colon cancer cells by antisense RNA and oligodeoxynucleotides. *Oncogene* 9, 291-298 (1994)

32. H. Huang, J.M. Trujillo & S. Chakrabarty: Proliferation of human colon cancer cells: role of epidermal growth factor and transforming growth factor. *Int J Cancer* 52, 978-986 (1992)

33. S. Chakrabarty, S. Rajagopal & S. Huang: Expression of antisense epidermal growth factor receptor RNA downmodulated the malignant behavior of human colon cancer cells. *Clin Exp Metastasis* 13, 191-195 (1995)

34. S. Rajagopal, S. Huang, T.L. Moskal, B-N. Lee, E.L. El-Naggar, & S.Chakrabarty: Epidermal growth factor expression in human colon and colon carcinomas: Antisense epidermal growth factor receptor RNA downregulates the proliferation of human colon cancer cells. *Int J Cancer* 62, 661-667 (1995)

35. R. Radinsky, S. Risin, D. Fan, Z. Dong, D. Bielenberg, C.D. Bucana & I.J. Fidler: Level and function of epidermal growth factor receptor predict the metastatic potential of human colon carcinoma cells. *Clin Cancer Res* 1, 19-31 (1995)

36. S. Chakrabarty, M.G. Brattain, R.L. Ochs & J. Varani: Modulation of fibronectin, laminin and cellular adhesion in

the transformation and differentiation of murine AKR fibroblasts. *J Cellular Physiol* 133, 415-425 (1987)

37. J. Varani & S. Chakrabarty: Modulation of fibronectin synthesis and fibronectin binding during transformation and differentiation of mouse AKR fibroblasts. *J Cellular Physiol* 143, 445-454 (1990)

38. J. Varani & S. Chakrabarty: Changes in the extracellular matrix during transformation and differentiation. In: *Microcirculation in cancer metastasis*. Eds: Orr F.W, Buchanan M.R, Weiss L, CRC Press, Boca Raton (1991)

39. D.C. Dean, T.M. Birkenmeier, C.D. Rosen, & S.J. Weintraub: Glycoprotein synthesis and secretion: expression of fibronectin and its cell surface receptors. *Am Rev Respir Dis* 144, S25-S28 (1991)

40. P.J. Wirth, L.D. Lup, Y. Fujimoto & H.C. Bisgaard: 1992. Two-dimensional electrophoretic analysis of transformation-sensitive polypeptide during chemically, spontaneously, and oncogene-induced transformation of rat liver epithelial cells. *Electrophoresis* 13, 305-320 (1992)

41. M.A. Schwartz: Spreading of human endothelial cells on fibronectin or vitronectin triggers elevation of intracellular free calcium. *J Cell Biol* 120, 1003-1010 (1993)

42. H.P. McNamee, D.E. Ingber & M.A. Schwartz: Adhesion to fibronectin stimulates inositol lipid synthesis and enhances PDGF-induced inositol lipid breakdown. *J Cell Biol* 121, 673-678 (1993)

43. K. Vuori and E. Ruoslahti: Activation of protein kinase C precedes  $\alpha 5 \beta 1$  integrin-mediated cell spreading on fibronectin. *J Biol Chem* 268, 21459-21462 (1993)

44. Z. Zhang, K. Vuori, J.C. Reed and E. Ruoslahti: The  $\alpha 5 \beta 1$  integrin supports survival of cells on fibronectin and up-regulates Bcl-2 expression. *Proc Natl Acad Sci (USA)* 92, 6161-6165 (1995)

45. D.G. Tang, M. Tarrien, P. Dobrzynski and K.V. Honn: Melanoma cell spreading on fibronectin induced by 12(S)-HETE involves both protein kinase C-and protein tyrosine kinase-dependent focal adhesion formation and tyrosine phosphorylation of focal adhesion kinase (pp125<sup>FAK</sup>). *J Cell Physiol* 165, 291-306 (1995)

46. F.G. Giancotti, and E. Ruoslahti: Elevated levels of the  $\alpha 5 \beta 1$  fibronectin receptor suppress the transformed phenotype of Chinese hamster ovary cells. *Cell* 60, 849-859 (1990)

47. C. Schreiner, M. Fisher, S. Hussein & R.L. Juliano: Increase tumorigenicity of fibronectin receptor deficient Chinese hamster ovary cell variants. *Cancer Res* 51, 1738-1740 (1991)

## Growth factors and colon cancer

48. D.M. Steel & H. Harris: The effect of antisense RNA to fibronectin on the malignancy of hybrids between melanoma and normal fibroblasts. *J Cell Sci* 93, 515-524 (1989)
49. S. Huang, J. Varani & S. Chakrabarty: Control of AKR fibroblast phenotype by fibronectin: regulation of cell-surface fibronectin binding receptor by fibronectin. *J Cell Physiol* 161, 470-482 (1994)
50. J. Varani, L. Schuger, S.E.G. Fligiel, D.R. Inman & S. Chakrabarty: Production of fibronectin by human tumor cells and interaction with exogenous fibronectin: comparison of cell lines obtained from colon adenocarcinomas and squamous carcinomas of the upper aerodigestive tract. *Int J Cancer* 47, 421-425 (1991)
51. H. Takei, Y. Iino, J. Horiguchi & T. Yokoe: Immunohistochemical fibronectin staining pattern and prognosis in invasive breast carcinoma. *Oncology* 52, 106-111 (1995)
52. S. Chakrabarty, S. Rajagopal, N.M. Navone & H.A. Fritsche: Potential of fibronectin as a new tumor marker for prostate cancer. *J Clin Ligand Assay* 20, 161 (1997)
53. S. Rajagopal, N.M. Navone, P. Troncoso, H.A. Fritsche & S. Chakrabarty: Characterization of the responses of a new human prostate carcinoma cell line MDA PCA2a to epidermal growth factor (EGF), transforming growth factor (TGF) and TGFbeta1. *Proc Am Assoc Cancer Res* 38, 554 (1997)
54. W.K.L. Chung: The role of stromal-epithelial interaction in normal and malignant growth. *Cancer Surveys* 23, 33-42 (1995)
55. H. Sonmez, S. Suer, L. Karaaslan, H. Baloglu & E. Kokoglu: Tissue fibronectin levels of human prostate cancer, as a tumor marker. *Cancer Biochem Biophys* 15, 107-110 (1995)
56. S. Suer, H. Sonmez, I. Karaaslan, H. Baloglu & E. Kokoglu: Tissue sialic acid and fibronectin levels in human prostate cancer. *Cancer Letters* 99, 135-137 (1996)
57. Clinical applications of TGF beta. Ciba foundation symposium 157. Eds: Bock G.R., Marsh J, John Wiley & Sons, NY (1991).
58. S. Cheifetz, J.A. Weatherbee, M.L-S. Tsang, J.K. Anderson, J.E. Mole, R. Lucas & J. Massague: The transforming growth factor-beta system, a complex pattern of cross-reactive ligands and receptors. *Cell* 48, 409-415 (1987)
59. K.A. Piez & M.B. Sporn: Transforming growth factor-beta chemistry, biology and therapeutics. *Annals NY Acad Sciences* 593, (1990)
60. J. Massague: Receptors for the TGFbeta family. *Cell* 69, 1067-1070 (1992)
61. L. Attisano, J. Carcamo, F. Ventura, F.M.B. Weis, J. Massague & J.L. Wrana: Identification of human activin and TGF beta type I receptors that form heteromeric kinase complexes with type II receptors. *Cell* 75, 671-680 (1993)
62. A. Moustakas, H.Y. Lin, Y.I. Henis, J. Plamondon, M.D. O'Connor-McCourt & H.F. Lodish: The transforming growth factor beta receptors types I, II, and III form hetero-oligomeric complexes in the presence of ligand. *J Biol Chem* 268, 22215-22218 (1993)
63. R. Ebner, R-H. Chen, L. Shum, S. Lawler, T.F. Zioncheck, A. Lee, A.R. Lopez & R. Derynck: Cloning of a type I TGF-beta receptor and its effect on TGF-beta binding to the type II receptor. *Science* (Washington, D.C.) 260, 1344-1348 (1993)
64. R.H. Chen, R. Ebner & R. Derynck: Inactivation of the type II receptor reveals two receptor pathways for the diverse TGFbeta activities. *Science* (Washington, D.C.) 260, 1335-1338 (1993)
65. J.L. Wrana, L. Attisano, R. Wieser, F. Ventura & J. Massague: Mechanism of activation of the TGFbeta receptor. *Nature* (London) 370, 341-346 (1994)
66. P.H. Howe, C.C. Bascom, M.R. Cunningham & E.B. Leof: Regulation of transforming growth factor beta1 action by multiple transducing pathways: evidence for both G protein-dependent and -independent signaling. *Cancer Res* 49, 6024-6031 (1989)
67. U.S. Murthy, M.A. Anzano, J.M. Stadel, & R. Greig: Coupling of TGFbeta-induced mitogenesis to G-protein activation in AKR-2B cells. *Biochem. Biophys Res Commun* 152, 1228-1235 (1988)
68. P.H. Howe & E.B. Leof: Transforming growth factor beta1 treatment of AKR-2B cells is coupled through a pertussis-toxin-sensitive G-protein(s). *Biochem J* 261, 879-886 (1989)
69. S. Chakrabarty: The role of protein kinase C in transforming growth factor beta1 induction of carcinoembryonic antigen in human colon carcinoma cells. *J Cell Physiol* 152, 494-499 (1992)
70. S. Chakrabarty & S. Huang: Role of protein kinase C in the induction of carcinoembryonic antigen by transforming growth factor beta1. *J Cell Physiol* 164, 148-153 (1995)
71. Y. Geng & R.A. Weinberg: Transforming growth factor beta effects on expression of G1 cyclins and cyclin-dependent protein kinases. *Proc Natl Acad Sci* (USA) 90, 10315-10319 (1993)
72. M.E. Kadin, M.W. Cavaille-Coll, R. Gertz, J. Massague, R. Kataoka, J. Sherlock & S.M. Lanier: Signalling events initiated by transforming growth factor beta1 that require G<sub>i</sub> 1. *J Biol Chem* 268, 19851-19857 (1993)



## Growth factors and colon cancer

73. M.G. Brattain, A.E. Levine, S. Chakrabarty, L.C. Yeoman, J.K.V. Willson & B. Long: Heterogeneity of human colon carcinoma. *Cancer Met Revs* 3, 177-191 (1984)
74. D.F. Pierce Jr., A.E. Gorska, A. Chytil, K.S. Meise, D.L. Page, R.J. Coffey & H.L. Moses: Mammary tumor suppression by transforming growth factor beta1 transgene expression. *Proc Natl Acad Sci (USA)* 92, 4254-4258 (1995)
75. L. Sun, G. Wu, J.K.V. Willson, E. Zborowska, J. Yang, I. Rajkarunanayake, J. Wang, L.E. Gentry, X-F. Wang & M.G. Brattain: Expression of transforming growth factor beta type II receptor leads to reduced malignancy in human breast MCF-7 cells. *J Biol Chem* 42, 26449-26455 (1994)
76. Y. Nishizuka: Intracellular signaling by hydrolysis of phospholipids and activation of protein kinase C. *Science* (Washington, D.C.) 258, 607-614 (1992)
77. C.A. O'Brian & N.E. Ward: Biology of the protein kinase C family. *Cancer Met Rev* 8, 199-214 (1989)
78. P.L. Baron, M.J. Koretz, R.A. Carchman, J.M. Collins, A.S. Tokarz & G.A. Parker: Induction of the expression of differentiation-related antigens on human colon carcinoma cells by stimulating protein kinase C. *Arch Surg* 125, 344-350 (1990)
79. G.C. Blobe, L.M. Obeid & Y.A. Hannun: Regulation of protein kinase C and role in cancer biology. *Cancer Met Rev* 13, 411-431 (1994)
80. L. Coussens, P.J. Parker, L. Rhee, T.L. Yang-Feng, E. Chen, M.D. Waterfield, U. Francke & A. Ullrich: Multiple, distinct form of bovine and human protein kinase C suggest diversity in cellular signaling pathways. *Science* (Washington, D.C.) 233, 859-866 (1986)
81. J.L. Knopf, M.H. Lee, L.A. Sultzman, R.W. Kriz, C.R. Loomis, R.M. Hewick & R.M. Bell: Cloning and expression of multiple protein kinase C cDNAs. *Cell* 46, 491-502 (1986)
82. P. Tremble, R. Chiquet-Ehrismann & Z. Werb: The extracellular matrix ligands fibronectin and tenascin collaborate in regulating collagenase gene expression in fibroblasts. *Mol Biol of the Cell* 5, 439-453 (1994)
83. D. Wang, T.M. Birkenmeier, J. Yang, S. Venkateswarlu, L. Humphrey, M.G. Brattain & L. Sun: Release from quiescence stimulates the expression of integrin 5beta1 which regulates DNA synthesis in human fibrosarcoma HT1080 cells. *J Cell Physiol* 164, 499-508 (1995)
84. S. Rajagopal, S. Huang, M. Albitar & S. Chakrabarty: Control of fibronectin receptor expression by fibronectin: antisense fibronectin RNA downmodulates the induction of fibronectin receptor by transforming growth factor beta1. *J Cell Physiol* 170, 138-144 (1997)
85. S. Reynolds, S. Ragagopal & S. Chakrabarty: Mechanisms of action of differentiation-inducing agents in human colon cancer cells. *Proc Am Assoc Cancer Res* 38, 498 (1997)
86. Y. Nishizuka: The role of protein kinase C in cell surface signal transduction and tumour promotion. *Nature* (London) 308, 693-698 (1984)
87. L. Liao, K. Ramsay & S. Jaken: Protein kinase C isozymes in progressively transformed rat embryo fibroblasts. *Cell Growth and Diff* 5, 1185-1194 (1994)
88. M-H. Disatnik, A.R. Winnier, D. Mochly-Rosen, & C.L. Arteaga: Distinct responses of protein kinase C isozymes to c-erbB-2 activation in SKBR-3 human breast carcinoma cells. *Cell Growth and Differen* 5, 873-880 (1994)
89. B. Liu, R.J. Maher, Y.A. Hannun, A.T. Porter & K.V. Honn: 12(S)-HETE enhancement of prostate tumor cell invasion: selective role of PKC. *J Natl Cancer Inst* 86, 1145-1151 (1994)
90. S. Ahmad, J.B. Trepel, S. Ohno, K. Suzuki, T. Tsuruo & R.I. Glazer: Role of protein kinase C in the modulation of multidrug resistance: expression of the atypical gamma isoform of protein kinase C does not confer increased resistance to doxorubicin. *Mol Pharmacol* 42,1004-9 (1992)
91. G.C. Blobe, C.W. Sachs, W.A. Khan, D. Fabbro, S. Stabel, W.C. Wetsel, L.M. Obeid, R.L. Fine & Y.A. Hannun: Selective regulation of expression of protein kinase C (PKC) isozymes in multidrug-resistant MCF-7 cells. *J Biol Chem* 268, 658-64 (1993)
92. H.H. Grunicke & F. Uberall: Protein kinase C modulation. *Seminars in Cancer Biol* 3, 351-60 (1992)
93. C.A. O'Brian, D. Fan, N.E. Ward, Z. Dong, Z. L. Iwamoto, K.P. Gupta, L.E. Earnest & I.J. Fidler: Transient enhancement of multidrug resistance by the bile acid deoxycholate in murine fibrosarcoma cells in vitro. *Biochem Pharmacol* 41, 797-806 (1991)
94. K.R. Gravitt, N.E. Ward, D. Fan, J.M. Skibber, B. Levin & C.A. O'Brian: Evidence that protein kinase C activation is a critical event in phorbol ester-induced multiple drug resistance in human colon cancer cells. *Biochem Pharm* 48, 375-381 (1994)
95. S. Chakrabarty & S. Huang: Modulation of chemosensitivity in human colon carcinoma cells by downregulating protein kinase C expression. *J Exp Therapeutics & Oncol* 1, 218-221 (1996)
96. M.A. Anzano, D. Rieman, W. Prichett, D.F. Bowen-Pope & R. Greig: Growth factor production by human colon carcinoma cell lines. *Cancer Res* 49, 2898-2904 (1989)

## Growth factors and colon cancer

97. T.A. Libermann, H.R. Nusbaum, N. Razon, R. Kris, I. Lax, H. Soreq, N. Whittle, M.D. Waterfield, A. Ullrich & J. Schlessinger: Amplification, enhanced expression and possible rearrangement of EGF receptor gene in primary human brain tumours of glial origin. *Nature* (London) 313, 144-147 (1985)
98. D.F. Stern, D.L. Hare, M.A. Cecchini & R.A. Weinberg: Construction of a novel oncogene based on synthetic sequences encoding epidermal growth factor. *Science* (Washington, D.C.), 235, 321-324 (1987)
99. F. Ciardiello, N. Kim, T. Saeki, R. Dono, M.G. Persico, G.D. Plowman, J. Garriques, S. Radke, G.J. Todaro & D.S. Salomon: Differential expression of epidermal growth factor-related proteins in human colorectal tumors. *Proc Natl Acad Sci* (USA) 88, 7792-7796 (1991)
100. R.J. Coffey, C.M. McCutchen, R. Graves-Deal & W.H. Polk Jr: Transforming growth factors and related peptides in gastrointestinal neoplasia. *J Cell Biochem Sppl* 16G, 111-118 (1992)
101. K. Khazaie, V. Schirmacher & R.B. Lichtner: EGF receptor in neoplasia and metastasis. *Cancer Metast Rev* 12, 155-274 (1993)
102. P. Borlinghaus, S. Wieser & R. Lamerz: Epidermal growth factor, transforming growth factor- $\alpha$ , and epidermal growth factor receptor content in normal and carcinomatous gastric and colonic tissue. *Clin Invest* 71, 903-907 (1993)
103. R.B. Myers, J.E. Kudlow & W.E. Grizzle: Expression of transforming growth factor- $\alpha$ , epidermal growth factor and epidermal growth factor receptor in adenocarcinoma of the prostate and benign prostatic hyperplasia. *Modern Pathology* 6, 733-737 (1993)
104. K.C. Ching, E. Ramsey, N.D. Pettigrew, R. Cunha, M. Jason & J.G. Dodd: Expression of mRNA for epidermal growth factor, transforming growth factor- $\alpha$  and their receptor in human prostate tissue and cell lines. *Mol Cell Biochem* 126, 151-158 (1993)
105. P. Limonta, R.M. Moretti, D. Dondi, M.M. Marelli & M. Motta: Androgen-dependent prostatic tumors: biosynthesis and possible actions of LHRH. *J Steroid Biochem and Mol Biol* 49, 347-350 (1994)
106. X.H. Liu, H.S. Wiley & A.W. Meikle: Androgens regulate proliferation of human prostate cancer cells in culture by increasing transforming growth factor- $\alpha$  (TGF- $\alpha$ ) and epidermal growth factor (EGF)/TGF- $\alpha$  receptor. *J Clin Endocrin & Metabol* 77, 1472-1478 (1993)
107. C.J. Fong, E.R. Sherwood, J. Mendelsohn, C. Lee & J.M. Kozlowski: Epidermal growth factor receptor monoclonal antibody inhibits constitutive receptor phosphorylation, reduces autonomous growth, and sensitizes androgen-independent prostatic carcinoma cells to tumor necrosis factor alpha. *Cancer Res* 52, 5887-5892 (1992)
108. R. Lupu, M. Cardillo, L. Harris, M. Hijazi & K. Rosenberg: Interaction between erbB-receptors and heregulin in breast cancer tumor progression and drug resistance. *Seminars in Cancer Biol* 6, 135-145 (1995)
109. L. Zhang, C-J Chang, S.S. Bacus & M-C Hung: Suppressed transformation and induced differentiation of HER-2/neu-overexpressing breast cancer cells by modemin. *Cancer Res* 55, 3890-3896 (1995)
110. S.A. Chrysogelos, R.I. Yarden, A.H. Lauber & J.M. Murphy: Mechanisms of EGF receptor regulation in breast cancer cells. *Breast Cancer Res & Treatment* 31, 227-236 (1994)
111. C.F. LeMaistre, C. Meneghetti, L. Howes & C.K. Osborne: Targeting the EGF receptor in breast cancer treatment. *Breast Cancer Res & Treatment* 32, 97-103 (1994)
112. L.H. Hartwell, & M.B. Kastan: Cell cycle control and cancer. *Science* (Washington, D.C.) 266, 1821-1827 (1994)
113. D. Peng, Z. Fan, Y. Lu, T. DeBlasio, H. Scher & J. Mendelsohn: Anti-epidermal growth factor receptor monoclonal antibody 225 up-regulates p27<sup>KIP1</sup> and induces G1 arrest in prostate cancer cell line DU145. *Cancer Res* 56, 3666-3669 (1996)
114. X. Wu, M. Rubin, Z. Fan, T. DeBlasio, T. Soos, A. Koff & J. Mendelsohn: Involvement of p27KIP1 in G1 arrest mediated by an anti-epidermal growth factor monoclonal antibody. *Oncogene* 12, 1397-403 (1996)
115. M.B. Datto, Y. Li, J.F. Panus Y. Xiong & X.F. Wang: Transforming growth factor beta induces the cyclin-dependent kinase inhibitor p21 through a p53-independent mechanism. *Proc Natl Acad Sci* (USA) 92, 5545-5549 (1995)
116. C.Y. Li, L. Suardet & J.B. Little: Potential role of WAF1/Cip1/p21 as a mediator of TGF- $\beta$  cytoinhibitory effect. *J Biol Chem* 270, 4971-4974 (1995)
117. A. Richardson, & J.T. Parsons: Signal transduction through integrins: a central role for focal adhesion kinase? *BioEssays* 17, 229-236 (1995)
118. E. Ruoslahti & J. Reed: Anchorage independence, integrins and apoptosis. *Cell* 77, 477-478 (1994)
119. M. Schaller & T. Parsons: pp125FAK-dependent tyrosine phosphorylation of paxillin creates a high affinity binding site for Crk. *Mol Cell Biol* 15, 2635-2645 (1995)
120. S.M. Frisch, K. Vuori, E. Rouslahti & P-Y Chan-Hui: Control of adhesion-dependent cell survival by focal adhesion kinase. *J Cell Biol* 134, 793-799 (1996)

## Growth factors and colon cancer

121. L.V. Owens, L. Xu, R.J. Craven, G.A. Dent, T.M. Weiner, L. Kornberg, E.T. Liu & W.G. Cance: Overexpression of focal adhesion kinase (p125 FAK) in invasive human tumors. *Cancer Res* 55, 2752-2755 (1995)
122. L. Xu, L.V. Owens, G.C. Sturge, X. Yang & E.T. Liu: Attenuation of the expression of the focal adhesion kinase induces apoptosis in tumor cells. *Cell Growth and Differentiation* 7, 413-418 (1996)
123. Antisense RNA and DNA. Current Communications in Molecular Biology Eds: Melton D.A., Cold Spring Harbor Laboratory (1988)
124. F. J-F. Liu: Serum tumor marker assays in cancer patient care. *The Cancer Bull U. T. M. D. Anderson Cancer Center* 45, 55-63 (1993)
125. J. Thompson & W. Zimmermann: The carcinoembryonic gene family: structure, expression and evolution. *Tumor Biol* 9, 63-83 (1988)
126. J.A. Thompson: Molecular cloning and expression of carcinoembryonic antigen gene family members. *Tumor Biol* 16, 10-16 (1995)
127. T. Osaki, Y. Tanio, I. Tachibana, H. Hosoe, T. Kumagai, I. Kawase, S. Oikawa & T. Kishimoto: Gene therapy for carcinoembryonic antigen-producing human lung cancer cells by cell type-specific expression of herpes simplex virus thymidine kinase gene. *Cancer Res* 54, 5258-5261 (1994)
128. T. Tanaka, F. Kanai, S. Okabe, Y. Yoshida, H. Wakimoto, H. Hamada, Y. Shirotori, K. Lan & M. Ishitobi: Adenovirus-mediated prodrug gene therapy for carcinoembryonic antigen producing human gastric carcinoma cells in vitro. *Cancer Res* 56, 1341-1345 (1996)
129. N. Zhu, D. Liggitt, Y. Liu & R. Debs: Systemic gene expression after DNA delivery into adult mice. *Science* (Washington, D.C.), 261, 209-211 (1993)
130. D. Yu, A. Matin, W. Xia, F. Sorgi, L. Huang & M.C. Hung: Liposome-mediated in vivo E1A gene transfer suppressed dissemination of ovarian cancer cells that overexpress Her-2/neu. *Oncogene* 11, 1383-1388 (1995)
131. X. Xing, A. Matin, D. Yu, W. Xia, F. Sorgi, L. Huang & M.C. Hung: Mutant SV40 large T antigen as a therapeutic agent for HER-2/neu-overexpressing ovarian cancer. *Cancer Gene Therapy* 3, 168-174 (1996)